<u>Indian J Clin Biochem</u>. 2013 Oct; 28(4): 314–328.

Published online 2013 Sep 1. doi: <u>10.1007/s12291-013-0375-3</u>

PMCID: PMC3783921

Vitamin C in Disease Prevention and Cure: An Overview

Shailja Chambial, Shailendra Dwivedi, Kamla Kant Shukla, Placheril J. John, and Praveen Sharma

Author information ► Article notes ► Copyright and License information ► This article has been <u>cited by</u> other articles in PMC. Go to:

Abstract

The recognition of vitamin C is associated with a history of an unrelenting search for the cause of the ancient haemorrhagic disease scurvy. Isolated in 1928, vitamin C is essential for the development and maintenance of connective tissues. It plays an important role in bone formation, wound healing and the maintenance of healthy gums. Vitamin C plays an important role in a number of metabolic functions including the activation of the B vitamin, folic acid, the conversion of cholesterol to bile acids and the conversion of the amino acid, tryptophan, to the neurotransmitter, serotonin. It is an antioxidant that protects body from free radical damage. It is used as therapeutic agent in many diseases and disorders. Vitamin C protects the immune system, reduces the severity of allergic reactions and helps to fight off infections. However the significance and beneficial effect of vitamin C in respect to human disease such as cancer, atherosclerosis, diabetes, neurodegenerative disease and metal toxicity however remains equivocal. Thus further continuous uninterrupted efforts may open new vistas to understand its significance in disease management.

Keywords: Vitamin C, Atherosclerosis, Diabetes, Immunity, Cancer, Infertility, Heavy metal toxicity

<u>Go to:</u>

Introduction

Vitamins are essential nutrients that are required for various biochemical and physiological processes in the body. It is well known that most of the vitamins cannot be synthesized in the body and hence their supplementation in diet is essential. Vitamins are classified on the basis of their solubility as water soluble (C and B complexes) and fat soluble vitamins (A, D, E, K). Vitamin C or ascorbic acid (AA) was first isolated in 1923 by Hungarian biochemist and Nobel laureate Szent-Gyorgyi and synthesized by Howarth and Hirst [1]. It exists in reduced [ascorbate] and oxidized forms as dehydroascorbic acid which are easily inter-convertible and biologically active thus it acts as important antioxidant. Vitamin C is easily oxidized acid and destroyed by oxygen, alkali and high temperature. Most of the plant and animal species have the ability to synthesize vitamin C from glucose and galactose through uronic acid pathway but man and other primates cannot do so because of deficiency of enzyme gulonolactone oxidase [EC 1.1.3.8] required for it's biosynthesis. Deficiency of this enzyme is a result of a mutation which occurred approximately 40 million years ago [2].

The body requires vitamin C for normal physiological functions. It helps in the synthesis and metabolism of tyrosine, folic acid and tryptophan, hydroxylation of glycine, proline, lysine

carnitine and catecholamine. It facilitates the conversion of cholesterol into bile acids and hence lowers blood cholesterol levels. It also increases the absorption of iron in the gut by reducing ferric to ferrous state. As an antioxidant, it protects the body from various deleterious effects of free radicals, pollutants and toxins. The therapeutic effect of vitamin C was explored by Linus Pauling however his work on therapeutic role of vitamin C in his later years generated much controversy yet he was the first to introduce the concept of high doses of vitamin C for the treatment of various conditions from common cold to cancer [3]. Since then mega doses of vitamin C have been widely used in the treatment and prevention of a large number of disorders like diabetes, atherosclerosis, common cold, cataracts, glaucoma, macular degeneration, stroke, heart diseases, cancer and so on.

Deficiency of this vitamin is often associated with anemia, infections, bleeding gums, scurvy, poor wound healing, capillary haemorrhage, muscle degeneration, atherosclerotic plaques and neurotic disturbances. For the correction of deficiency, vitamin C is often supplemented in large doses and unlike fat soluble vitamins, toxicity is rare. Recently the role of vitamin C in infection and immunity has also been investigated. In view of the vast biological, physiological functions and therapeutic role of vitamin C, this review is an attempt to summarise various evidences in this context.

Go to:

Dietary Sources of Vitamin C

Vitamin C is found in citrus fruits, green peppers, red peppers, strawberries, tomatoes, broccoli, brussels sprouts, turnip, Indian gooseberry and other leafy vegetables. The animal sources are poor in vitamin C content and the level is usually <30–40 mg/100 g. Therefore plant sources become important because of high content of vitamin C up to 5,000 mg/100 g. It's absorption in the buccal cavity is by passive diffusion however in gastrointestinal tract absorption is by active sodium dependent vitamin C transporters (SVCT) [4, 5].

Go to:

Vitamin C Bioavailability

Bioavailability or the effective concentration of vitamin C essentially depends on its effective absorption from intestine and renal excretion. Vitamin C, consumed either with diet or dietary supplements, is absorbed by the epithelial cells of the small intestine by SVCT1 or, subsequently diffuses into the surrounding capillaries and then the circulatory system [5–7]. Circulating AA is filtered from kidney capillary bed into the Bowman's capsule through a general filtration mechanism. AA is reabsorbed through SVCT1 transporter in proximal convoluted tubule [6]. The difference between the amount of AA filtered and reabsorbed constitutes renal excretion [8]. Together, intestinal absorption and renal excretion controls the serum level of vitamin C and thus its bioavailability. At low concentrations, most vitamin C is absorbed in the small intestine and reabsorbed from the renal tubule [9]. However, at high concentrations, SVCT1 is down regulated [10] which limits the amount of AA absorbed from the intestine and kidney [11]. This imposes a physiological restriction on the maximal effective serum vitamin C concentration (or its bioavailability) that is attainable by oral consumption [12]. This value has been determined to be 200 mmol/L [12], although "normal" physiological serum concentrations of ascorbate in healthy humans range from 60 to

100 mmol/L [13]. Vitamin C levels in circulating blood cells, such as platelets, are much higher than the plasma [13], as these cells express the SVCT2 transporter [14], which mediates intracellular ascorbate accumulation [15].

Reduced bioavailability of vitamin C is often seen in stress, alcohol intake, smoking, fever, viral illnesses, usage of antibiotics, pain killers, exposure to petroleum products or carbon monoxide, heavy metals toxicity and so on. However, the precise mechanism behind low vitamin C level in the body is not well defined. Presumably, an increased utilization of this vitamin in these conditions and/or a decreased absorption from the gut could be a cause of decreased vitamin C level [16]. In the event of high consumption, AA and its metabolites such as dehydroascorbic acid, 2,3-diketogulonic acid and oxalic acid are excreted via kidney in humans. AA is generally non-toxic but at high doses (2–6 g/day) it can cause gastrointestinal disturbances or diarrhoea [17, 18]. The side effects are generally not serious and can be easily reversed by reducing the intake of AA. Furthermore, there is no consistent and compelling data on serious health effects of vitamin C in humans [18, 19].

Go to:

Biochemical Functions of Vitamin C

The biochemical functions of AA are largely dependent on the oxido-reduction properties of 1-AA which is a co-factor for hydroxylation and activity of mono-oxygenase enzymes in the synthesis of collagen, carnitine and neurotransmitters [20]. AA accelerates hydroxylation reactions by maintaining the active centre of metal ions in a reduced state for optimal activity of enzymes hydroxylase and oxygenase. Thus, it is crucial in the maintenance of collagen which represents about one-third of the total body protein. In an experimental study AA has been shown to have involvement in synthesis and release of type IV collagen into the culture medium [21]. Further, it has also been reported that AA 2-phosphate, a long-acting vitamin C derivative, stimulates both cell growth and the expression of mRNA for type III collagen in human osteoblast-like MG-63 cells and in normal human osteoblasts, as well as in human bone marrow mesenchymal stem cells, but not the expression of type I collagen in these cells [22]. However, in another study Kishimoto et al. [23] have observed that AA induced the expression of type 1 and type 4 collagen and SVCT2 in cultured human skin fibroblasts. Collagen constitutes the principal protein of skin, bones, teeth, cartilage, tendons, blood vessels, heart valves, inter vertebral discs, cornea, eye lens. AA is essential to maintain the enzyme prolyl and lysyl hydroxylase in an active form. AA deficiency results in reduced hydroxylation of proline and lysine, thus affecting collagen synthesis. AA is also an essential co-factor for hydroxylations involved in the synthesis of muscle carnitine "β-hydroxybutyric acid" [24, 25]. Carnitine is required for the transport and transfer of long chain fatty acids into mitochondria for energy production. Further, AA is also a co-factor for the enzyme dopamineβ-hydroxylase, which catalyzes the conversion of neurotransmitter dopamine to norepinephrine [19] and hence essential for the synthesis of catecholamines. In addition, AA catalyzes other enzymatic reactions involving amidation necessary for maximal activity of hormones oxytocin, vasopressin, cholecystokinin and alpha-melanotropin [26]. It is also involved in the transformation of cholesterol to bile acids as it modulates the microsomal 7αhydroxylation, the rate limiting reaction of cholesterol catabolism in liver [27]. Deficiency of AA affects this conversion and as a result cholesterol accumulates in the liver leading to hypercholesterolemia [28, 29], cholesterol gall stones formation etc [30, 31].

Go to:

Vitamin C and Common Cold

Apart from the well accepted role of vitamin C in the prevention of scurvy, the most widely known health beneficial effect of AA is in the prevention and relief of common cold. Pauling was the first to introduce the concept of high dose of vitamin C and suggested that ingestion of 1–3 g of AA effectively prevents/ameliorates common cold [32]. The role of oral vitamin C in the prevention and treatment of cold however remains controversial despite many controlled studies [33]. A number of clinical trials with varying doses of AA showed that it does not have significant prophylactic effect, but reduces the severity and duration of symptoms of cold during the period of infection. Randomized and non-randomized trials on role of vitamin C in prevention and treatment of the common cold indicated that AA in dose of 1 g/day during severe winter months produced no beneficial effect on the incidences of common cold [34]. In both preventive and therapeutic trials, there was a consistent beneficial but generally modest therapeutic effect on the duration of cold symptoms. There was no clear indication of the relative benefits of different regimes of vitamin C doses. However, in trials that tested vitamin C after cold symptoms occurred, there was some evidence of greater benefits with larger doses than with lower doses [34, 35]. Attenuation of immunity in common cold is well known. There has been a continuous debate about the role of AA in boosting immunity during rhinitis. AA has been shown to stimulate immune system by enhancing T-cell proliferation in response to infection. These cells are capable of lysing infected targets by producing large quantities of cytokines and by helping B cells to synthesize immunoglobulins to control inflammatory reactions. Further, it has been shown that AA blocks pathways that lead to apoptosis of T-cells and thus stimulate or maintain Tcell proliferation to attack the infection. This mechanism has been proposed for the enhanced immune response observed after administration of vitamin C during rhinitis [36, 37].

Go to:

Vitamin C and Tissue Healing

It is also fairly understood that wound healing requires synthesis and accumulation of collagen and subsequent cross-linking of the fibre to give new tensile strength to the damaged tissue. An early study demonstrated that maximum tensile strength of scar tissue in guinea pig was achieved after supplementation of vitamin C [38]. Since then various studies have been carried out to evaluate the role of AA in wound repair and healing/regeneration process as it stimulates collagen synthesis. Adequate supplies of AA are necessary for normal healing process especially for post-operative patients because there is rapid utilization of AA for the synthesis of collagen at the site of wound/burns during post-operative period hence, administration of 500 mg to 1.0 g/day of AA are recommended to accelerate the healing process [39]. Of late, Jagetia et al. [40] demonstrated that AA pre-treatment was beneficial in healing of irradiated wounds and suggested a vitamin C related therapeutic strategy to accelerate wound repair in such conditions and in the cases of combined injury situation.

Go to:

Vitamin C and Iron

AA is known to enhance the availability and absorption of iron from non-heme iron sources [41]. It's supplementation is found to facilitate the dietary absorption of iron. The reduction of

iron by AA has been suggested to increase dietary absorption of non-heme iron [42, 43]. Vitamin C rich fruits such as goose berry has been reported to increase the bioavailability of iron from staple cereals and pulses [44]. Recent observations are of the view that vitamin C inhibits the expression of hepcidin and by affecting erythropoietin receptor in HepG2 cells and the bioavailability of iron provides protection against anaemia due to iron deficiency [45]. Darius Lane et al. [46], has considered ascorbate as a novel modulator for the classical transferrin Fe⁺ uptake pathway, acting through intracellular reductive mechanism. It is also well known that AA acts as a pro-oxidant in invitro in the presence of redox-active iron and might contribute to the formation of hydroxyl radical, which eventually may lead to lipid, DNA or protein oxidation [47]. Thus vitamin C supplementation in individuals with high iron and or bleomycin detectable iron in some preterm infants could be deleterious due to the production of oxidatively damaged molecules [48–51]. However, Proteggente et al. [52] have observed no pro-oxidant effect of AA supplementation on DNA damage in presence or absence of iron.

Go to:

Vitamin C and Fertility

Vitamin C has been used in the management of male infertility on empirical grounds, particularly in the presence of non-specific seminal infections [53]. It's presence in the seminal plasma of healthy adults in high concentration, ranging from 2.5 to 12 mg/dL, has been reported by various authors [54, 55]. However, the precise role of vitamin C in relation to male reproduction is not yet clear. Chinoy [56] stated that AA was essential for the structural and functional integrity of androgen-dependent reproductive organs. Low concentration of vitamin C showed marked degenerative changes in the testes, epididymis and vas deferens of scorbutic guinea pigs [57]. Besides degeneration of the spermatogenic epithelium, steroidogenesis and plasma testosterone level also showed a decline [58]. On the other hand, excessive intake of vitamin C has been reported to cause reproductive failure in the males [59]. However, Sapra et al. [60] could not observe any definite effect of vitamin C supplementation on Leydig cells in guinea pigs. AA as the principle antioxidant in seminal plasma of fertile men, contributes up to 65 % of its total chain breaking antioxidant capacity [61]. It's concentration in seminal plasma is almost ten times higher than plasma concentration. In various studies AA content in seminal plasma of fertile and infertile men was found to be significantly different [62, 63] and the percentage of sperm with normal morphology correlated significantly with seminal AA in both the groups [63–65]. AA deficiency may lead to an increase in oxidative damage induced by reactive oxygen species (ROS) and increased ROS was observed in the semen of 25–45 % of infertile men [66]. This is further corroborated by association of decreased AA with increase seminal plasma lipid peroxidation as observed in human trial [67]. Others have also observed oxidative stress induced deleterious effect on male fertility [68, 69]. Increased free radicals in the seminal plasma of infertile subjects by lowering the effective vitamin C levels may potentiate the deleterious effects that result in abnormal sperm parameters [70, 71]. Further studies report that supplementation of AA leads to significant reduction in ROS concentration [72, 73], sperm membrane lipid per-oxidation [73] and sperm DNA oxidation [74] and increased sperm quality [72–74]. The results of a recent animal experimental study indicated that, vitamin C improves antioxidant enzymes activity and significantly reduce MDA in testis compared with the test group [75]. Vitamin C supplementation as antioxidant in dose dependent manner in men may improve sperm quality [76]. It's supplementation also increases progesterone levels in infertile women with luteal phase defect [77].

Vitamin C and Atherosclerosis

There are several publications on the role of vitamin C in lipid metabolism and atherogenesis with diverse observations. The significance of dietary inadequacy of vitamin C in the aetiology of dyslipidemia and atherosclerosis first became apparent from the clinical studies of Myasnikova in 1947. The study showed lowering of cholesterol level by administration of AA in the hypercholesterolemic patients [78]. Since then several authors have also persuaded similar studies. One reviewed the evidence for the role of vitamin C in bile acid synthesis [27] while others gave particular emphasis on the potential involvement of vitamin C in pathogenesis of atherosclerosis [79, 80]. There are reports indicating increase in total body cholesterol and hypercholesterolemia in acutely scorbutic guinea pigs. However, some studies could not observe any effect of vitamin C in similar animal models [81]. Das et al. [82] observed that administration of AA lowers blood cholesterol, triglycerides, lipid per-oxidation and increases HDL cholesterol. Most of the earlier studies were conducted using rabbit as an animal model for examining vitamin C deficiency. As such rabbit is not a suitable model for such studies as it can synthesize AA unlike higher primates and human beings. It is difficult to elicit vitamin C deficiency in animal models. In view of the conflicting observations based on the acutely scorbutic animal model chosen by most of the workers, Ginter et al. [83] designed a model of chronic latent vitamin C deficiency in guinea pigs. This model in contrast to others, enabled the effect of AA deficiency on lipid metabolism and atherosclerosis to be followed in long term experiments. In protracted hypovitaminosis C lasting for 10 weeks, there was a considerable accumulation of cholesterol in liver and also increased concentration in serum [83–85].

It was also reported that the deficiency of vitamin C leads to enhanced accumulation of cholesterol in thoracic aorta along with pathomorphological changes in blood vessels [83, 86, 87]. Various human trials have also indicated vitamin C induced reduction in blood lipid levels in normal and hypercholesterolemic subjects [88, 89]. Marc and Kothari and Sharma have further observed that vitamin C administration causes significant reduction in LDL and increase in HDL [87] and there by provides protection against CAD [87, 90]. Similar observations have been given by others also [91–96]. Chronic AA deficiency in man can lead to impaired cholesterol metabolism resulting in atheromatous changes in the vascular system [87]. This is further supported by the observation that vitamin C lowers cholesterol [88] and reduces the risk of developing cardiovascular disease (CHD) [97, 98]. Numerous studies have also looked into the association between AA intake and blood lipids. A large prospective epidemiological study in Finnish men and women suggested that high intake of AA was associated with a reduced risk of death from CHD in women than in men [98]. Similarly, several other studies showed that high intake of AA in American men and women appeared to benefit only women [97, 99]. Yet, another cohort study suggested that cardiovascular mortality was reduced in both sexes by vitamin C [100]. It is likely that cholesterol lowering effect of vitamin C is affected by several factors like initial cholesterol levels, age and sex of the subjects, dose and mode of the administration. The influence of age may be important because SAA levels have been found to be lower in elderly as compared to adolescents [101, 102] and therefore elderly subjects could be more responsive to the administration of vitamin C. In UK, a study showed that the risk of stroke in those with highest intake of vitamin C was only half that of subjects with the lowest intake. No evidence is suggestive of lower rate of CHD in those with high vitamin C intake [103]. A recent meta-analysis study on the role of AA and antioxidant vitamins also showed no evidence of significant benefit in prevention of

CHD [104]. Thus, no conclusive evidence is available on the possible protective effect of AA supplementation on CHD.

Increased attention is being paid to involvement of low density lipoprotein (LDL) in atherogenesis. There are reports indicating that lipid peroxidation and oxidative modification of LDL are implicated in development of atherosclerosis [105]. Vitamin C provides protection against oxidative changes in LDL in different types of oxidative stress including metal induced oxidative stress [106]. Addition of iron to plasma devoid of AA resulted in lipid peroxidation, whereas endogenous and exogenous AA was found to inhibit the lipid oxidation in iron-over loaded human plasma [107]. In an invitro study, when AA was added to human serum supplemented with Cu²⁺, antioxidant activity were observed rather than pro-oxidant effects [108]. AA is known as important antioxidant that scavenges free radicals and thus primarily prevents the oxidation of LDL in aqueous medium [109]. In addition, invitro studies have also shown that AA strongly inhibits LDL oxidation by vascular endothelial cells at physiological concentrations [110–112]. An important factor that initiates atherosclerosis is the adhesion of leukocytes to the endothelium. Invivo studies have shown that AA inhibits leukocyte-endothelial cell interactions induced by cigarette smoke [113, 114] or oxidized LDL [115]. Further, lipophilic derivatives of AA showed protective effect on lipid-peroxide induced endothelial injury [116]. In endothelial cells, AA prevented atherogenic modification of mildly oxidized LDL [110] and preserved α -tocopherol in both cells and LDL [117]. Although AA may not reverse established atherosclerosis, it can prevent the endothelial dysfunction that is the earliest sign of many vascular inflammatory conditions. AA is responsible for increased endothelial cell proliferation and also checks tumour necrosis factor (TNF) alpha induced endothelial cell growth inhibition. Vitamin C as antioxidant helps in endothelial cell proliferation and also correlated with expression of collagen IV in endothelial cells. Study has also shown that when proliferating endothelial cells were treated with AA, increased retinoblastoma protein (Rb) phosphorylation was observed with decreased level of p53 as compared to untreated cells. Furthermore, the addition of AA to TNF-alpha-treated proliferating endothelial cells blocked both the inhibition of retinoblastoma protein phosphorylation and enhanced p53 expression induced by TNF-alpha and TNF-alpha-induced apoptosis [117].

A number of studies were carried out in humans to determine the protective effect of AA supplementation (500–100 mg/day) on invivo and exvivo lipid peroxidation in healthy individuals and smokers with inconclusive findings. AA supplementation showed reduction or no change in lipid peroxidation products [111, 118-122]. There are reports where vitamin C has been found to decrease LDL peroxidation even in passive smokers [123]. In this context, it is important to note that during exvivo LDL oxidation, AA is removed at initial stage of LDL isolation from plasma therefore, no change in exvivo appears [124]. May and Li, examined the role of vitamin C in oxidation of LDL which causes endothelial dysfunction, an important early manifestation of atherosclerosis. They observed that up-regulation of endothelial cell SVCT2 expression and function may help to maintain intracellular ascorbate during oxLDL-induced oxidative stress, and that ascorbate in turn can prevent this effect [125]. Overall, both invitro and invivo experiments showed that AA protects isolated LDL and plasma lipid peroxidation induced by various radicals or oxidants generating systems. However, there are reports in experimental animals that large doses of exogenous iron and AA promote the release of iron from iron binding proteins and also enhance invitro lipid peroxidation in serum. This finding supports the hypothesis that high intake of iron along with AA could increase invivo lipid peroxidation of LDL and therefore could increase risk of atherosclerosis [126]. However, Chen et al. [127], demonstrated that ascorbic acts as an antioxidant towards lipids even in presence of iron load invivo systems. Vitamin C also helps in prevention of atherosclerosis by strengthening the artery walls through its participation in the synthesis of collagen and by preventing the undesirable adhesion of white blood cells to damaged arteries [128–132].

Go to:

Vitamin C and Cancer

The notion that vitamin C may have a preventive role in cancer was first proposed in 1949. It was demonstrated by Cameron et al. [133–135], that high-dose vitamin C improved the survival of patients with terminal cancer. However, the first documented study in which vitamin C was administered to cancer patients was carried out in the 1970s, by Pauling and Cameron. They gave 10 g (10,000 mg) of vitamin C per day to 100 terminally ill cancer patients and compared their outcome with 1,000 cancer patients who were given conventional therapy. It was observed that 10.3 % cancer patients receiving vitamin C survived while all patients on conventional therapy without vitamin C died [134]. Other studies have also confirmed these findings. Murata and Morishige showed in a study conducted on Japanese patients with uterus cancer receiving 5–30 g of vitamin C that these patients survived six times longer than those on vitamin C <4 g per day. When comparison was made between those supplemented with or without vitamin C, survival rate was 15 % higher in those supplemented with vitamin C [136]. The overwhelming evidence supports that a high intake of vitamin C is linked with a low risk for cancer of oesophagus, oral cavity, stomach, pancreas, cervix, rectum and breast [137, 138] and also non-hormonal cancers [139]. One of the most important amenable determinants of cancer risk is diet. Several research panels and committees have independently concluded that high fruit and vegetable intake reduces the risk of different types of cancer [140, 141] and mortality rate was also found to be inversely related to vitamin C intake [142, 143]. However a study involving 34,000 post-menopausal women, reported no such association between the intake of vitamins A, C and E and a reduced risk of developing breast cancer [144]. Intravenous vitamin C has also been reported to have beneficial effect in advanced cancer [145]. Several mechanisms proposed indicating involvement of vitamin C in the treatment and prevention of cancer are: enhancing the immune system; stimulating the formation of collagen; preventing metastasis (spreading) by inhibiting enzymes; preventing viruses that can cause cancer; correction of vitamin C deficiency which is often associated with cancer patients; wound healing in cancer patients after surgery; enhancing the effectiveness of chemotherapy; reducing the toxicity of chemotherapy; preventing free radical damages and neutralising some carcinogens [146].

Recently a number of experimental studies have observed that different types of cancer cells either do not grow at high vitamin C concentration or it leads to tumour shrinkage [147, 148]. Further recent experimental studies have also found that ascorbate supplementation hinders metastasis, tumour growth and inflammatory cytokine secretion as well as enhanced encapsulation of tumours in Gulo KO mice [149, 150]. Reports have shown that intravenous injection increases vitamin C concentration more than 70 times in relation to oral administration and effectiveness of treatment is linked to vitamin C concentration [12, 145]. Thus controversy is because of mode, dose and duration of administration.

Newly available pharmacokinetic data improved the understanding of the regulation of vitamin C transport, and the growing evidence on the therapeutic efficacy of vitamin C. This has stimulated interest to reassess the feasibility of using vitamin C in the prevention and

treatment of cancer. Though different in their methodologies, most recent studies on vitamin C and cancer have two central themes:(1) the effects of high-dose AA on the development and progression of tumours; and (2) the mechanisms of action that may contribute to the anticancer effect [144]. Research has also refocused on the implications and applicability of high i.v. dose of vitamin C in cancer therapy. In contrast to normal physiological concentration of AA (0.1 mmol/L) pharmacological concentrations of AA (0.3–20 mmol/L) selectively targets and kills tumour cells in invitro. This tumour-killing phenomenon is attributable to the prooxidant property of vitamin C, which, at high concentration mediates the production of hydrogen peroxide thus provides a potential mechanism of action for the anti-tumour effect of vitamin C and it's implication as a pro-drug in cancer treatment [145, 147]. However, it is difficult to assess the precise contribution of vitamin C in the clinical outcome, as subjects under examination simultaneously receive different therapeutic treatments [151]. Therefore, the therapeutic value of high-dose vitamin C administration in cancer progression or remission is not unequivocally supported but i.v. administration of vitamin C in high doses improves the health-related quality of life even at the advanced stage of the disease [152].

Go to:

Vitamin C and Diabetes

Diabetes is becoming a pandemic and numbers are expected to rise to 366 million (4.4 % of the global population) by 2030 [153]. In diabetic patients, long-term damage, dysfunction, and failure of different organs, especially the eyes (diabetic retinopathy), kidneys (diabetic nephropathy), nerves (diabetic neuropathy), heart (myocardial infarction), and blood vessels (atherosclerosis) are related to uncontrolled hyperglycaemia [154–156]. Hyperglycaemia induces oxidative stress [157] primarily by ROS [18]. There is convincing experimental and clinical evidence that the generation of ROS increases in both types of diabetes and that the onset of diabetes is closely associated with oxidative stress [158]. Vitamin C has been associated with decreased risk of developing diabetes mellitus (DM). In Norfolk Prospective Study the association between fruit and vegetable intake and plasma levels of vitamin C and risk of type 2 DM was established [159]. During 12-years of follow-up, 735 incident cases of diabetes were identified among nearly 21,000 participants. A significant inverse association was found between plasma levels of vitamin C and risk of diabetes (odds ratio = 0.38, 95 % confidence interval: 0.28–0.52) [159]. This is further supported by a study with longer follow up of 23 years which reported that antioxidants induced risk reduction of type 2 diabetes [160] and vitamin C level was found to be significantly lower in both insulin dependent and non dependent diabetes [161, 162]. Vitamin C reduces fasting and postprandial oxidative stress [163]. Sharma et al. have observed reduced vitamin C levels in diabetic subjects. They further reported that vitamin C level is also associated with various components of metabolic syndrome and with the increment in component there is a sharp reduction in vitamin C level [164]. In recent experimental studies it has been found that Vitamin C and E supplementation relieves oxidative stress in the blood and tissues of diabetic aged rats by modulating the antioxidant system and lipid profile [165, 166].

Diabetes is associated with various micro vascular and macro vascular complications. Hyperglycaemia in diabetes is responsible for micro vascular ROS generation which causes endothelial dysfunction [167] and vitamin C blocks acute hyperglycaemic impairment of endothelial function in diabetic subjects [168]. One of the most important micro vascular complications is diabetic nephropathy. According to statistical prediction, out of 30 million patients with diabetes in India, diabetic nephropathy is expected to develop in 6.66 million

[169]. Qin et al. [170] reported that vitamin C supplementation significantly decreased podocyte injury in diabetic rats. Perhaps AA protects podocyte by increasing antioxidative capacity and ameliorating the renal oxidative stress [171]. The role of vitamin C in diabetic retinopathy has also been reported in various studies. Vitamin C and E supplementation reduces neovascularization, prevent the inhibition of retinal glutathione reductase, glutathione peroxidase and superoxide dismutase activities; hence vitamin C and E prevent oxidative stress induced retinopathy [172–174]. Neuropathy is also one of the micro vascular complications often manifested in uncontrolled DM. Some studies report that the role of vitamin C in diabetic neuropathy is not as well pronounced as other antioxidants [175]. Some suggest that the AA levels are significantly low in diabetic polyneuropathy patients [176]. Role of vitamin C and other dietary antioxidants have been reviewed by several authors with controversial findings [177].

Go to:

Vitamin C and Immunity

Vitamin C affects several components of the human immune system. Vitamin C appears to play a role in a number of neutrophil functions including increased chemotaxis, increased particulate ingestion, enhanced lysozyme-mediated non-oxidative killing, protection against the toxic effects of superoxide anion radical, inhibition of the halide-peroxidemyeloperoxidase system without a pronounced bactericidal effect, and stimulation of the hexose monophosphate shunt [178].

The role of vitamin C seems to be more pronounced in cell mediated response instead of humoral immunity as the T-cell hyporesponsiveness was observed to be reversed in Crohn's disease patients on oral supplementation of vitamin C. In the same study no effect was observed on humoral immunity [179]. Another study supports the fact that vitamin C acts with other micronutrients synergistically and enhances skin barrier function as well as protective activities of immune cells but its role in antibody protection is not as pronounced [180]. On the contrary animal studies support the role of supplementation of vitamin C in humoral immunity as it increases serum levels of antibodies [181] and C1q complement proteins [182] in guinea pigs, which cannot synthesize vitamin C like humans and hence depend on dietary supplementation. Vitamin C along with other micronutrients help in reverting potential damage caused by free radicals at cellular level and modulates immune cell functions through regulation of redox-sensitive transcription factors and affects production of cytokines and prostaglandins. Adequate intake of vitamins C along with other vitamins and micronutrients like B₆, folate, B₁₂, E, selenium, zinc, copper, and iron supports a Th1 cytokine-mediated immune response [183, 184] with sufficient production of proinflammatory cytokines, which maintain an effective immune response. Supplementation with these micronutrients reverses the Th2 cell-mediated immune response to Th1 cytokineregulated response with enhanced innate immunity [183].

Vitamin C inhibits the excessive activation of the immune system to prevent tissue damage. It also supports antibacterial activity, stimulates natural killer (NK) cells and differentiation of Th0 subset into Th1 subset [184, 185]. In addition, vitamin C also modulates synthesis of proinflammatory cytokines, or expression of adhesive molecules [185].

Mikirova et al. [186] have demonstrated that intravenous vitamin C treatment reduces proinflammatory cytokines IL-1 α , IL-2, IL-8, TNF- α , chemokine eotaxin and CRP in cancer

patients. Several studies have shown that the modulation of inflammation by intravenous vitamin C correlated with decrease in tumour marker levels [186–188]. Studies conducted on human subjects reported that that plasma vitamin C and dietary intakes of vitamin C are inversely associated with some markers of the acute phase response and haemostasis that have been associated with greater risk of CVD and non-vascular disease. Plasma vitamin C, fruit intake, and dietary vitamin C intake were significantly and inversely associated with mean concentrations of C-reactive protein, an acute phase reactant, and tissue plasminogen activator antigen, a marker of endothelial dysfunction, even after adjustment for confounders. The findings suggest that vitamin C has anti-inflammatory effects and is associated with lower endothelial dysfunction in men with no history of CHD or diabetes [189–196]. Vitamin C concentrations in the plasma and leukocytes rapidly decline during infections and stress. Supplementation of vitamin C has been shown to improve components of the human immune system such as antimicrobial and NK cell activities, lymphocyte proliferation, chemotaxis, and delayed-type hypersensitivity as discussed above. Vitamin C contributes in maintenance of the redox integrity of cells and thereby protects them against ROS generated during the respiratory burst and in the inflammatory response [197].

Thus, vitamin C has diverse role as an antioxidant protecting the immune cells against intracellular ROS production during inflammatory response, acting as an enzymatic cofactor and maintaining tissue integrity and plays a crucial role in formation of skin, epithelial and endothelial barriers [185]. Of late vitamin C supplementation has been found to be beneficial in various inflammatory conditions.

Go to:

Vitamin C and Heavy Metal Toxicity

Metals including iron, copper, chromium, and vanadium undergo redox cycling, while cadmium, mercury, and nickel, as well as lead deplete glutathione and protein-bound sulfhydryl groups, resulting in the production of ROS as superoxide ion, hydrogen peroxide, and hydroxyl radical. As a consequence, enhanced lipid peroxidation, DNA damage, and altered calcium and sulfhydryl homeostasis occur [198]. Various experimental studies report the beneficial effect of vitamin C against heavy metal toxicity. Lead is considered as one of the common environmental poison in which protective role of vitamin C is extensively studied. A recent experimental study based on histopathological examination revealed the diminution of detrimental effects of chronic lead intoxication on liver, kidneys, brain and testes [199]. In another study lead induced electrophysiological changes were inhibited in rat colon by AA administration [200]. The beneficial effect of AA on lead concentrations in human studies is however inconclusive. A large survey comprising of 19,578 participants (6–90 years) without prior history of lead poisoning reported that blood levels of AA are inversely related to plasma AA and recent dietary intake had no influence on blood levels. This study surmises that there may be a protective relationship between AA and lead [201].

Arsenic toxicity is essentially associated with lipid peroxidation and oxidative stress. Arsenic in drinking water may even cause chromosomal aberration leading to molecular disorders [202]. Arsenic exposure during gestation and lactation leads to significantly increased lipid peroxidation in the rat brain which was reversed by supplementation of vitamin C, E and Zn [203]. In another study arsenic induced normocytic and normochromic anemia as well as a significant increase in hemolysis, TBARS production, catalase activity, hyperlipidemia, and impairment in renal functions in mice pups during gestation and lactation which was partially

reverted by the administration of AA [204]. Arsenic induced hepatotoxicity has also been reported by recent experimental studies which have suggested that vitamin C supplementation improves mitochondrial structure and function along with restriction of apoptosis due to caspase-3 inhibition in arsenic trioxide exposed rat liver. The overall report is of view that vitamin C and vitamin C rich fruits such as goose berry provides protection against metal induced hepatotoxicity [205, 206].

Cadmium is an extremely toxic metal commonly found in industrial workplaces similar to lead and arsenic also causes lipid peroxidative changes in various tissues. An experimental study discussed protective role of vitamin C supplementation in lung and brain of rat exposed to excessive cadmium [207]. Vitamin C also reverted haematological changes in mercury and cadmium exposed Wistar rats [208]. Vitamin C was also observed to be protective against concomitant exposure to heavy metal and radiation in another experimental study [209].

Go to:

Vitamin C and Neurodegenerative Disorders

Schizophrenia is one of the major neurological disorders associated with great deal of morbidity and economic burden. It is a multifactorial disease and hence has a poor outcome in spite of the best available treatments. It is worth mentioning that simple water soluble vitamin C, adequately present in fruits and vegetables had drawn attention of the psychiatrists almost seven decades ago for the treatment of schizophrenia. A study conducted on 12 schizophrenics showed that urinary excretion of vitamin C was significantly lower than healthy controls and intravenous injection of large dose of vitamin C produced improvement in mental condition in 75 % of patients [210]. In another study it was observed that vitamin C level was significantly low in plasma and urine of schizophrenics as compared to normal controls. Administration of vitamin C improved plasma vitamin C level and thus concluded that schizophrenic patients require higher levels of vitamin C than the suggested optimal AA requirement for healthy individuals [211]. Several investigators have implicated role of increased free radical generation in pathogenesis of schizophrenia. Alteration in the optimum activities of antioxidant enzymes [212–214] and related parameters of lipid peroxidation [215, 216] in blood have been detected in schizophrenics. Brain contains large amount of unsaturated fatty acids, catecholamines and monoamines, which are the target molecules for lipid peroxidation [217, 218]. Brain is rich in iron containing compounds and thus it is an easy target for lipid peroxidation through the formation of hydroxyl radicals. Monoamine and catecholamine oxidation also produces superoxide anions in the brain [219]. AA, an antioxidant vitamin, plays an important role in protecting free radical-induced damage in the brain. Dadheech et al. [220] reported the antioxidant deficit in schizophrenics and it was associated with increased MDA level in blood which is a marker for lipid peroxidation. Vitamin C is present in dopamine dominant areas at high concentrations in the brain tissue as compared to other organs [221, 222]. Recently, Arvindakshan et al. [223] reported reduction in brief psychiatric rating scale (BPRS) and positive and negative syndrome scale score after supplementation with omega-3 fatty acids, vitamin C, and vitamin E. A decrease in the levels of tocopherol, total AA and reduced glutathione was found in schizophrenics compared to normal controls. Further a significant rise in oxidative stress and decreased antioxidant status was observed in the chronic stage of schizophrenia as compared to those in acute condition. A significant rise in dehydroascorbic acid with concomitant fall in reduced AA suggests scavenging action of AA and its utilization with increased oxidative stress as indicated by high blood malondialdehyde levels. Leucocyte AA, a better index of AA status was also

found to be reduced in schizophrenics, suggesting depletion of body stores of AA and the condition worsened with advancing age [224].

Very few studies have examined the effect of vitamin C with typical antipsychotics in the treatment of schizophrenia. Oral supplementation of vitamin C with antipsychotic reverses AA levels, reduces oxidative stress, and improves BPRS score, hence both the drugs in combination can be used in the treatment of schizophrenia [225]. The findings of another study suggest that antioxidant supplement therapy as an adjuvant therapy is useful in patients with stress-induced psychiatric disorders [226]. There are also reports advocating beneficial effect of vitamin C in neurodegenerative disorders including Alzheimer's disease. Overall there is large body of evidence supporting that maintaining healthy vitamin C level can have a protective function against age related cognitive decline but avoiding vitamin C deficiency is likely to be more beneficial than taking supplements on top of normal healthy diet [227].

Go to:

References

- 1. Haworth WN, Hirst EL. Synthesis of ascorbic acid. J Soc Chem Ind (Lond) 1933;52:645–647.
- 2. Nishikimi M, Fukuyama R, Minoshima S, Shimizu N, Yagi K. Cloning and chromosomal mapping of the human nonfunctional gene for l-gulono-gamma-lactone oxidase, the enzyme for l-ascorbic acid biosynthesis missing in man. J Biol Chem. 1994;269:13685–13688. [PubMed]
- 3. Dunitz JD. Linus Carl Pauling. 28 February 1901–19 August 1994. Biographical Memoirs of Fellows of the Royal Society. 1996;42:316–8. [PubMed]
- 4. Stevenson NR, Brush MK. Existence and characteristic of Na⁺ dependent active transport of ascorbic acid in guinea pigs. Am J Clin Nutr. 1969;22:318. [PubMed]
- 5. Malo C, Wilson JX. Glucose modulates vitamin C transport in adult human small intestinal brush border membrane vesicles. J Nutr. 2000;130:63–69. [PubMed]
- 6. Takanga H, Mackenzie B, Hediger MA. Sodium dependent ascorbic acid transporter family SLC23. Pflugers Arch. 2004;447:677–682. [PubMed]
- 7. Stewart JS, Booth CC. Ascorbic acid absorption in malabsorption. Acta Gastroenterol Belg. 1964;27:567–568. [PubMed]
- 8. Ralli EP, Friedman GJ, Rubin SH. The mechanism of the excretion of vitamin C by the human kidney. J Clin Invest. 1938;17:765–770. [PMC free article] [PubMed]
- 9. Nelson EW, Lane H, Fabri PJ, Scott B. Demonstration of saturation kinetics in the intestinal absorption of vitamin C in man and the guinea pig. J Clin Pharmacol. 1978;18:325–335. [PubMed]
- 10. MacDonald L, Thumser AE, Sharp P. Decreased expression of the vitamin C transporter SVCT1 by ascorbic acid in a human intestinal epithelial cell line. Br J Nutr. 2002;87(20):97–100. [PubMed]
- 11. Wilson JX. Regulation of vitamin C transport. Ann Rev Nutr. 2005;25:105–125. [PubMed]
- 12. Padayatty SJ, Sun H, Wang YH, Riordan HD, Hewitt SM, Katz A, et al. Vitamin C pharmacokinetics: implications for oral and intravenous use. Ann Intern Med. 2004;140:533–537. [PubMed]
- 13. Levine M, Conry-Cantilena C, Wang Y, Welch RW, Washko PW, Dhariwal KR, et al. Vitamin C pharmacokinetics in healthy volunteers: evidence for a recommended dietary allowance. Proc Natl Acad Sci USA. 1996;93:3704–3709. [PMC free article] [PubMed]

- 14. Dixon SJ, Wilson JX. Adaptive regulation of ascorbate transport in osteoblastic cells. J Bone Miner Res. 1992;7:675–681. [PubMed]
- 15. Li Y, Schellhorn Herb E. New developments and novel therapeutic perspectives for vitamin C. J Nutr. 2007;137:2171–2184. [PubMed]
- 16. Kallner AB, Hartmann D, Horning DH. Steady state turnover and body pool of ascorbic acid in man. Am J Clin Nutr. 1979;32:530–539. [PubMed]
- 17. Anderson D, Phillips BJ, Yu T, Edwards AJ, Ayesh R, Butterworth KR. The effect of vitamin C supplementation on biomarkers of oxygen radical generated damage in human volunteers with low or high cholesterol levels. Environ Mol Mutagen. 1997;30:161–174. [PubMed]
- 18. Johnson CS. Biomarkers for establishing a tolerable upper intake level for vitamin C. Nutr Rev. 1999;57:71–77. [PubMed]
- 19. Naidu AK, Vitamin C. In human health and disease is still a mystery? An overview. Nutr J. 2003;2:7. [PMC free article] [PubMed]
- 20. Levin M. New concepts in the biology and biochemistry of ascorbic acid. New Engl J Med. 1986;31:892–902. [PubMed]
- 21. May JM, Zhi-chao Q. Transport and intracellular accumulation of vitamin C in endothelial cells: relevance to collagen synthesis. Arch Biochem Biophys. 2005;434:178–186. [PubMed]
- 22. Maehata Y, Takamizawa S, Ozawa S, Izukuri K, Kato Y, Sato S, et al. Type III collagen is essential for growth acceleration of human osteoblastic cells by ascorbic acid 2-phosphate, a long-acting vitamin C derivative. Matrix Biol. 2007;26(5):371–381. [PubMed]
- 23. Kishimoto Y, Saito N, Kurita K, Shimokado K, Maruyama N, Ishigami A. Ascorbic acid enhances the expression of type 1 and type 4 collagen and SVCT2 in cultured human skin fibroblasts. Biochem Biophys Res Commun. 2013;30(2):579–584. [PubMed]
- 24. Hulse JD, Ellis SR, Henderson LM. Carnitine biosynthesis-beta hydroxylation of trimethyllysine by an α -ketoglutarate dependent mitochondrial dioxygenase. J Biol Chem. 1978;253:1654–1659. [PubMed]
- 25. Rebouche CJ. Ascorbic acid and carnitine biosynthesis. Am J Clin Nutr. 1991;54(6):1147S–1152S. [PubMed]
- 26. Cameron E, Pauling L. Ascorbic acid and the glycosaminoglycans. Oncology. 1973;27:181–192. [PubMed]
- 27. Anon. Ascorbic acid and catabolism of cholesterol. Nutr Rev 1973;31:154. [PubMed]
- 28. Sharma P, Pramod J, Kothari LK, Ranka R, Sharma S. Hyperlipidemia in guinea pigs induced by chronic vitamin C deficiency. IJCB. 1989;4:62–64.
- 29. Sharma P, Pramod J, Sharma PK, Sapra M, Kothari LK. Effect of vitamin C deficiency and excess on the liver: a histopathological and biochemical study in guinea pigs fed normal or high cholesterol diet. Ind J Pathol Microbiol. 1990;33(4):307–313. [PubMed]
- 30. Ginter E, Bobek P, Jurcovicova M. Role of ascorbic acid in lipid metabolism. In: Seith PA, Toblert BM, editors. Ascorbic acid, chemistry, metabolism and uses. Washington DC: American Chemical Society; 1982. pp. 381–393.
- 31. Gustafsson U, Wang FH, Axelson M, Kallner A, Sahlin S, Einarsson K. The effect of vitamin C in high doses on plasma and biliary lipid composition in patients with cholesterol gallstones: prolongation of the nucleation time. Eur J Clin Invest. 1997;27(5):387–391. [PubMed]
- 32. Pauling L. Vitamin C and common cold. San Francisco: Freeman; 1970.
- 33. Elwood PC, Lee HP, St Leger AS, Baird M, Howard AN. A randomized controlled trial of vitamin C in the prevention and amelioration of the common cold. Br J Prev Soc Med. 1976;30(3):193–196. [PMC free article] [PubMed]
- 34. Hemilä H, Chalker E. Vitamin C for preventing and treating the common cold. Cochrane Database Syst Rev. 2013;1:CD000980. doi:10.1002/14651858.CD000980. [PubMed]

- 35. Douglas RM, Chalker EB, Treacy B. Vitamin C for preventing and treating the common cold. Cochrane Database Syst Rev. 2000;2:CD000980. [PubMed]
- 36. Campbell JD, Cole M, Bunditrutavorn B, Vell AT. Ascorbic acid is a potent inhibitor of various forms of T cell apoptosis. Cell Immunol. 1999;194:1–5. [PubMed]
- 37. Wintergerst ES, Maggini S, Hornig DH. Immune-enhancing role of vitamin C and zinc and effect on clinical conditions. Ann Nutr Metab. 2006;50(2):85–94. [PubMed]
- 38. Bourne GH. The effect of vitamin C on healing wounds. Proc Nutr Soc. 1946;4:204. [PubMed]
- 39. Hellman L, Burns JJ. Metabolism of l-ascorbic acid-1-C14 in man. J Biol Chem. 1958;230:923–930. [PubMed]
- 40. Jagetia GC, Rajanikant GK, Mallikarjun Rao KVN. Ascorbic acid increases healing of excision wounds of mice whole body exposed to different doses of γ-radiation. Burns. 2007;33(4):484–494. [PubMed]
- 41. Hallberg L. Bioavailability of dietary iron in man. Annu Rev Nutr. 1981;1:123–127. [PubMed]
- 42. Bendich A, Cohen M. Ascorbic acid safety: analysis factors affecting iron absorption. Toxicol Lett. 1990;51:189–190. [PubMed]
- 43. Zhang Y, Zhao D, Xu J, Xu C, Dong C, Liu Q, et al. Effects of dietary factors on the pharmacokinetics of (58)Fe-labeled hemin after oral administration in normal rats and the iron-deficient rats. Biol Trace Elem Res. 2013;153(1–3):243–250. [PubMed]
- 44. Gowri S, Patel K, Prakash J, Srinivasan K. Influence of amla fruits (*Emblica officinalis*) on the bio-availability of iron from staple cereals and pulses. Nutr Res. 2001;21(12):1483–1492.
- 45. Chiu PF, Ko SY, Chang CC. Vitamin C affects the expression of hepcidin and erythropoietin receptor in HepG2 cells. J Ren Nutr. 2012;22(3):373–376. [PubMed]
- 46. Darius Lane JR, Chikhani S, Richardson V, Richardson Des R. Transferrin iron uptake is stimulated by ascorbate via an intracellular reductive mechanism. Biochim Biophys Acta. 2013;1833(6):1527–1541. [PubMed]
- 47. Samuni A, Aronovitch J, Godinger D, Chevion M, Czapski G. On the cytotoxicity of vitamin C and metal ions: a site specific Fenton mechanism. Eur J Biochem. 1983;137:119–120. [PubMed]
- 48. Minetti M, Forte T, Soriani M, Quaresima V, Menditto A, Ferrari M. Iron Induced ascorbate oxidation in plasma as monitored by ascorbate free radical formation: no spin trapping evidence for the hydroxyl radical in iron-over loaded plasmas. Biochem J. 1992;282:459–465. [PMC free article] [PubMed]
- 49. Berger TM, Mumby S, Gutteridge JMC. Ferrous ion detected in iron-overloaded cord blood plasma from preterm and term babies: implication for oxidation stress. Free Radic Res. 1995;22:555–559. [PubMed]
- 50. Halliwell B. Vitamin C: antioxidant or pro-oxidant in vivo ? Free Radic Res. 1996;25:439–454. [PubMed]
- 51. Herbert V, Shaw S, Jayatileke E. Vitamin C driven free radicals generation from iron. J Nutr. 1996;126:1213–1220. [PubMed]
- 52. Proteggente AR, Rehman A, Halliwell B, Rice-Evans CA. Potential problems of ascorbic acid and iron supplementation: pro-oxidant effect in vivo? Biochem Biophys Res Commun. 2000;277:535–54053. [PubMed]
- 53. Mathur V, Murdia A, Hakim AA, Suhalka ML, Shaktawat GS, Kothari LK. Male infertility and the present status of its management by drugs. J Postgrad Med. 1979;25:90–96. [PubMed]
- 54. Huggins C, Scott WW, Heinen JH. Chemical composition of human serum and of the secretion of prostate and seminal vesicles. Am J Physiol. 1942;136:467–473.

- 55. Mann T. Biochemistry of semen and of the male reproductive tract. London: Methnen and Co. Ltd.; 1954. p. 20.
- 56. Chinoy NJ. Ascorbic acid turn over in animal and human tissue. J Anim Morphol Physiol. 1978; (Silver Jubilee Volume):69–85.
- 57. Chinoy MR, Sharma JD, Sanjeevan AG, Chinoy NJ. Structural changes in male reproductive organs and spermatozoa of scorbutic guinea-pigs. Proc Ind Natl Sci Acad. 1983;B49:628–635.
- 58. Gomes S, Odour OD, Bharaj B, Verjee ZH. Gonadal and plasma testosterone and cholesterol in scorbutic guinea-pigs. Int J Vit Nutr Res. 1977;47:75–80. [PubMed]
- 59. Paul PK, Datta-Gupta PN. Beneficial or harmful effects of a large dose of vitamin C on the reproductive organs of the male rat depending upon the level of food intake. Ind J Exp Biol. 1978;16:18–21. [PubMed]
- 60. Sapra M, Sharma P, Kothari LK. Effect of vitamin C deficiency on testicular structure in the guinea pig. J Postgrad Med. 1987;33:69–73. [PubMed]
- 61. Agarwal A. Role of antioxidants in treatment of male infertility. Reprod Biomed Online. 2004;8:616–627. [PubMed]
- 62. Abasalt H, Colagar Eisa T, Marzony Ascorbic acid in human seminal plasma: determination and its relationship to sperm quality. J Clin Biochem Nutr. 2009;45(2):144–149. [PMC free article] [PubMed]
- 63. Saleh RA, Agarwal A, Nada EA, Tonsy MH, Sharma RK, Meyer A, et al. Negative effects of increased sperm DNA damage in relation to seminal oxidative stress in men with idiopathic and male factor infertility. Fertil Steril. 2003;79:1597. [PubMed]
- 64. Shi YC, Sun HM, Shang XJ, Zhu PY, Huang YF. Total antioxidant capacity of seminal plasma in fertile and infertile men. Zhonghua Nan Ke Xue. 2005;11:915–917. [PubMed] 65. Shi YC, Shang XJ, Wang XL, Huang YF. Correlation of total antioxidant capacity in seminal plasma with sperm motility of infertile men. Zhonghua Nan Ke Xue. 2006;12:703–705. [PubMed]
- 66. Agarwal A, Ikemoto I, Loughlin KR. Relationship of sperm parameters to levels of reactive oxygen species in semen specimens. J Urol. 1994;152:107–110. [PubMed]
- 67. Shukla KK, Mahdi AA, Ahmad MK, Jaiswar SP, Shankwar SN, Tiwari SC. Mucuna pruriens reduces stress and improves the quality of semen in infertile men. Evid Based Complement Alternat Med. 2010;7(1):137–144. [PMC free article] [PubMed]
- 68. Hampl R, Drábková P, Kand'ár R, Stěpán J. Impact of oxidative stress on male infertility. Ceska Gynekol. 2012;77(3):241–245. [PubMed]
- 69. Doshi SB, Khullar K, Sharma RK, Agarwal A. Role of reactive nitrogen species in male infertility. Reprod Biol Endocrinol. 2012;15(10):109. [PMC free article] [PubMed]
- 70. Patriarca M, Menditto A, Morisi G. Determination of ascorbic acid in blood or serum and in seminal plasma using a simplified sample preparation and high performance liquid chromatography coupled with UV detection. J Liq Chromatogr. 1991;14:297–312.
- 71. Aitken RJ, Baker MA. Oxidative stress, sperm survival and fertility control. Mol Cell Endocrinol. 2006;250:66–69. [PubMed]
- 72. Donnelly ET, McClure N, Lewis SE. Glutathione and hypotaurine in vitro: effects on human sperm motility, DNA integrity and production of reactive oxygen species. Mutagenesis. 2000;15:61–68. [PubMed]
- 73. Kodama H, Yamaguchi R, Fukuda J, Kasi H, Tanaka K. Increased oxidative deoxyribonucleic acid damage in spermatozoa of infertile male patients. Fertil Steril. 1997;68:519–524. [PubMed]
- 74. Donnelly E, McClure N, Lewis SEM. The effect of ascorbate and alpha-tocopherol supplementation in vitro on DNA integrity and hydrogen peroxide induced DNA damage in human spermatozoa. Mutagenesis. 1999;14:505–512. [PubMed]

- 75. Jelodar G, Nazifi S, Akbari A. The prophylactic effect of vitamin C on induced oxidative stress in rat testis following exposure to 900 MHz radio frequency wave generated by a BTS antenna model. Electromagn Biol Med. 2013. doi:10.3109/15368378.2012.735208. [PubMed] 76. Verma A, Kanwar KC. Human sperm motility and lipid peroxidation in different ascorbic acid concentrations: an invitro analysis. Andrologia. 1998;30:325–329. [PubMed]
- 77. Henmi H, Endo T, Kitaiima Y, et al. Effects of ascorbic acid supplementation on serum progesterone levels in patients with a luteal phase defect. Fertil Steril. 2002;80:459–461. [PubMed]
- 78. Myasnikova IA. Effect of ascorbic acid, nicotinic acid and thiamine on cholestolemia. Voenno Morski Med Akad Leningrad. 1947;8:140.
- 79. Ginter E, Zolch Z. Raised ascorbic acid consumption in cholesterol fed guinea pigs. Int J Vit Nutr Res. 1972;42:72. [PubMed]
- 80. Krumdieck C, Butterworth CE. Ascorbate—cholesterol—lecithin interactions factors of potential importance in pathogenesis of atherosclerosis. Am J Clin Nutr. 1974;27:866. [PubMed]
- 81. Banerjee S, Gosh PK. Metabolism of acetate in scorbutic guinea pigs. Am J Physiol. 1960;199:1064. [PubMed]
- 82. Das S, Snehlata, Srivastava LK. Effect of ascorbic acid on lipid profile and per-oxidation in hypercholestrolemic rabbits. Nutr Res. 1997;17(2):231–241.
- 83. Ginter E, Babla J, Cerven J. The effect of chronic hypovitaminosis C on the metabolism of cholesterol and atherogenesis in guinea pigs. J Atheroscler Res. 1969;10:341. [PubMed]
- 84. Frikke-Schmidt H, Lykkesfeld J. Role of marginal vitamin C deficiency in atherogenesis: in vivo models and clinical studies. 2009. doi:10.1111/j.1742-7843.2009.00420. [PubMed]
- 85. Frikke-Schmidt H, Tveden-Nyborg P, Muusfeldt Birck M, Lykkesfeldt J. High dietary fat and cholesterol exacerbates chronic vitamin C deficiency in guinea pigs. Br J Nutr. 2011;105:54–61. [PubMed]
- 86. Sharma P, Pramod J, Sharma PK, Chaturvedi SK, Kothari LK. Effect of vitamin C administration on serum and aortic lipid profile of guinea pigs. IJMR. 1988;87:28. [PubMed] 87. Kothari LK, Sharma P. Aggravation of cholesterol induced hyperlipidemia by chronic vitamin C deficiency: experimental study in guinea pigs. Acta Biol Hung. 1988;39(1):4. [PubMed]
- 88. Vaney N, Sharma P, Pramod J, Varandami J, Kothari LK. Leucocyte ascorbic acid and blood lipids in normocholesterolemic men receiving different amounts of vitamin C. Vitaminologia. 1988;4:47–48.
- 89. Gaur GS, Dixit AK. Comparative study of vitamin C on serum lipid profile in healthy male and female human subjects. J Sci Res. 2012;4(3):775–781.
- 90. Marc P, McRae Vitamin C supplementation lowers serum low-density lipoprotein cholesterol and triglycerides: a meta-analysis of 13 randomized controlled trials. JCM. 2008;7(2):548–581. [PMC free article] [PubMed]
- 91. Itoh R, Yamada K, Oka J, Echizen H, Suyama Y, Murakami K. Serum ascorbic acid and HDL cholesterol in a healthy elderly Japanese population. Int J Vit Nutr Res. 1990;60(4):360–365. [PubMed]
- 92. Jacques PF. Effects of vitamin C on high-density lipoprotein cholesterol and blood pressure. J Am Coll Nutr. 1992;11(2):139–144. [PubMed]
- 93. Jacques PF, Sulsky SI, Perrone GA, Schaefer EJ. Ascorbic acid and plasma lipids. Epidemiology. 1994;5(1):19–26. [PubMed]
- 94. Jacques PF, Sulsky SI, Perrone GE, Jenner J, Schaefer EJ. Effect of vitamin C supplementation on lipoprotein cholesterol, apolipoprotein, and triglyceride concentrations. Ann Epidemiol. 1995;5(1):52–59. [PubMed]

- 95. Okamoto K. The relationship between dietary ascorbic acid intake and serum lipid concentration in the aged. Nihon Ronen Igakkai Zasshi. 1992;29(12):908–911. [PubMed] 96. Simon JA, Hudes ES. Relation of serum ascorbic acid to serum lipids and lipoproteins in US adults. J Am Coll Nutr. 1998;17(3):250–255. [PubMed]
- 97. Manson JE, Stampfer MJ, Willett WC, et al. A prospective study of vitamin C and incidence of coronary heart disease in women. Circulation. 1982;85:865–875.
- 98. Knekt P, Reunanen A, Jarvinen R, Seppanen R, Heliovaara M, Aromaa A. Antioxidant vitamin intake and coronary mortality in a longitudinal population study. Am J Epidemiol. 1994;139:1180–1189. [PubMed]
- 99. Rimm EB, Stampfer MJ, Ascherio A, Giovanno E, Colditz GA, Willettt WC. Vitamin E consumption and risk of coronary heart disease in men. N Engl J Med. 1993;328:1450–1456. [PubMed]
- 100. Enstrom JE, Kanim LE, Klein MA. Vitamin C intake and mortality among a sample of the United States population. Epidemiology. 1992;3:194–202. [PubMed]
- 101. Pramod J, Sharma P, Kothari LK. Effect of age and sex on serum and leucocyte ascorbic acid levels in normal human subjects. Vitaminologia. 1986;2:93–100.
- 102. Kothari LK, Pramod J, Sharma P, Chaturvedi SK. Influence of age and vitamin C status on serum cholesterol. IJE. 1988;17(4):929–930. [PubMed]
- 103. Gale CR, Martyn CN, Winter PD, Cooper C. Vitamin C and risk of death from stroke and coronary heart disease in cohort of elderly people. Br Med J. 1995;310:1563–1566. [PMC free article] [PubMed]
- 104. Ness A, Egger M, Davey-Smith G. Role of antioxidant vitamins in prevention of cardiovascular disease. Br Med J. 1999;319:577–579. [PMC free article] [PubMed]
- 105. Steinbrecher UP, Zhang H, Lougheed M. Role of oxidative modified LDL in atherosclerosis. Free Radic Biol Med. 1990;9:155–168. [PubMed]
- 106. Frei B. Vitamin C as an antiatherogen: mechanism of action. In: Packer L, Fuchs J, editors. Vitamin C in health and disease. New York: Marcel and Dekker Inc; 1997. pp. 163–182.
- 107. Berger TM, Polidori MC, Dabhag A, Evans PJ, Halliwell B, Marrow JD, et al. Antioxidant activity of vitamin C in iron over loaded human plasma. J Biol Chem. 1992;272:15656–15660. [PubMed]
- 108. Dasgupta A, Zdunek T. In vitro lipid peroxidation of human serum catalyzed by copper ion: antioxidant rather than prooxidant role of ascorbate. Life Sci. 1992;50:2875–2882. [PubMed]
- 109. Frei B, England L, Ames BN. Ascorbate is an outstanding antioxidant in human blood plasma. Proc Natl Acad Sci USA. 1989;86:6377–6381. [PMC free article] [PubMed]
- 110. Martin A, Frei B. Both intracellular and extracellular vitamin C inhibit atherogenic modification of LDL by human vascular endothelial cells. Atheroscler Thromb Vasc Biol. 1997;17:1583–1590. [PubMed]
- 111. Das S, Snehlata DN, Srivastava LM. Role of ascorbic acid on in vitro oxidation of low-density lipoprotein derived from hypercholesterolemic patients. Clin Chim Acta. 2006;372(1-2):202–205. [PubMed]
- 112. Shariat SZAS, Mostafavi SA, Khakpour F. Antioxidant effects of vitamins C and E on the low-density lipoprotein oxidation mediated by myeloperoxidase. Iran Biomed J. 2013;17(1):22–28. [PMC free article] [PubMed]
- 113. Lehr HA, Frei B, Arfors KE. Vitamin C prevents cigarette smoke-induced leukocyte aggregation and adhesion to endothelium in vivo. Proc Natl Acad Sci USA. 1994;91:7688–7692. [PMC free article] [PubMed]

- 114. Lehr HA, Weyrich AS, Saetzler RK, Jurek A, Arfors KE, Zimmerman GA, et al. Vitamin C blocks inflammatory platelet-activating factor mimetics created by cigarette smoking. J Clin Invest. 1997;99:2358–2364. [PMC free article] [PubMed]
- 115. Lehr HA, Frei B, Olofsson AM, Carew TE, Arfors KE. Protection from oxidized LDL induced leukocyte adhesion to microvascular and macrovascular endothelium in vivo by vitamin C but not by vitamin E. Circulation. 1995;91:1552–1553. [PubMed]
- 116. Kaneko T, Kaji K, Mastuo M. Protective effect of lipophilic derivatives of ascorbic acid on lipid peroxide-induced endothelial injury. Arch Biochem Biophys. 1993;304:176–180. [PubMed]
- 117. Sabharwal AK, May JM. Alpha-lipoic acid and ascorbate prevent LDL oxidation and oxidant stress in endothelial cells. Mol Cell Biochem. 2008;309:125–132. [PubMed]
- 118. Anderson D, Phillips BJ, Yu T, Edwards AJ, Ayesh R, Butterworth KR. The effect of vitamin C supplementation on biomarkers of oxygen radical generated damage in human volunteers with low or high cholesterol levels. Environ Mol Mutagen. 1997;30:161–174. [PubMed]
- 119. Fuller CJ, Grundy SM, Norkus EP, Jialal I. Effect ascorbate supplementation on low density lipoprotein oxidation in smokers. Atherosclerosis. 1996;119:139–150. [PubMed]
- 120. Nyyssonen K, Poulsen HE, Hayn M, Agerbo P, Porkkalo Sarataho E, Kaikkonen J, et al. Effect of supplementation of smoking men with plain or slow release ascorbic acid on lipoprotein oxidation. Eur J Clin Nutr. 1997;51:154–163. [PubMed]
- 121. Samman S, Brown AJ, Beltran C, Singh S. The effect of ascorbic acid on plasma lipids and oxidisability of LDL in male smokers. Eur J Clin Nutr. 1997;51:472–477. [PubMed]
- 122. Wen Y, Cooke T, Feely J. The effects of pharmacological supplementation with vitamin C on low density lipoprotein oxidation. Br J Clin Pharmacol. 1997;44:94–97. [PMC free article] [PubMed]
- 123. Valkonen MM, Kuusi T. Vitamin C prevents the acute atherogenic effects of passive smoking. Free Radic Biol Med. 2000;28(3):3–428. [PubMed]
- 124. Carr AC, Frei B. Does vitamin C act as pro-oxidant under physiological conditions? FASEB J. 1999;13:1007–24. [PubMed]
- 125. May JM, Li L, Qu ZC. Oxidized LDL up-regulates the ascorbic acid transporter SVCT2 in endothelial cells. ZC Mol Cell Biochem. 2010;343(12):217–222. [PMC free article] [PubMed]
- 126. Kapsokefalou M, Miller DD. Iron loading and large doses of intravenous ascorbic acid promote lipid peroxidation in whole serum in guinea pigs. Br J Nutr. 2001;85:681–687. [PubMed]
- 127. Chen K, Suh J, Carr AC, Marrow JD, Zeind J, Frei B. Vitamin C suppresses lipid damage in vivo even in the presence of iron over-load. Am J Physiol Endocrinol Metab. 2000;279:E1212–E1406. [PubMed]
- 128. Ginter E, Adama M. Characterization of the aortic collagens in guinea pigs with chronic vitamin C deficiency. Atherosclerosis. 1983;46(3):369–373. [PubMed]
- 129. Rath M. Eradicating heart disease. San Francisco: Health Now; 1993.
- 130. Lehr HA. Protection from oxidized LDL induced leukocyte adhesion to microvascular and macrovascular endothelium in-vivo by vitamin C but not by vitamin E. Circulation. 1995;91:1525–1532. [PubMed]
- 131. Weber C. Increased adhesiveness of isolated monocytes to endothelium is prevented by vitamin C intake in smokers. Circulation. 1996;93:1488–1492. [PubMed]
- 132. Nakata Y, Maeda N. Vulnerable atherosclerotic plaque morphology in apolipoprotein Edeficient mice unable to make ascorbic acid. Circulation. 2002;105:1485–1490. [PubMed]

- 133. Cameron E, Campbell A. The orthomolecular treatment of cancer. II. Clinical trial of high-dose ascorbic acid supplements in advanced human cancer. Chem Biol Interact. 1974;9:285–315. [PubMed]
- 134. Cameron E, Pauling L. Supplemental ascorbate in the supportive treatment of cancer: prolongation of survival times in terminal human cancer. Proc Natl Acad Sci USA. 1976;73:3685–3689. [PMC free article] [PubMed]
- 135. Cameron E, Pauling L. Supplemental ascorbate in the supportive treatment of cancer: reevaluation of prolongation of survival times in terminal human cancer. Proc Natl Acad Sci USA. 1978;75:4538–4542. [PMC free article] [PubMed]
- 136. Murata A, Morishige F, Yamaguchi H. Prolongation of survival times of terminal cancer patients by administration of large doses of ascorbate. Int J Vit Nutr Res. 1982;2/23:103–113. [PubMed]
- 137. Block G. Vitamin C and cancer prevention: the epidemiologic evidence. Am J Clin Nutr. 1991;53(1):270S–282S. [PubMed]
- 138. Block G. Epidemiologic evidence regarding vitamin C and cancer. Am J Clin Nutr. 1991;54(6):1310S. [PubMed]
- 139. Head KA. Ascorbic acid in the prevention and treatment of cancer. Altern Med Rev. 1998;3(3):174–186. [PubMed]
- 140. Block G, Patterson B, Subar A. Fruit, vegetables, and cancer prevention: a review of the epidemiological evidence. Nutr Cancer. 1992;18:1–29. [PubMed]
- 141. Steinmetz KA, Potter JD. Vegetables, fruit, and cancer prevention: a review. J Am Diet Assoc. 1996;96:1027–1039. [PubMed]
- 142. Loria CM, Klag MJ, Caulfield LE, Whelton PK. Vitamin C status and mortality in US adults. Am J Clin Nutr. 2000;72:139–145. [PubMed]
- 143. Khaw KT, Bingham S, Welch A, Luben R, Wareham N, Oakes S, Day N. Relation between plasma ascorbic acid and mortality in men and women in EPIC-Norfolk prospective study: a prospective population study: European prospective investigation into cancer and nutrition. Lancet. 2001;357:657–663. [PubMed]
- 144. Kushi LH, Fee RM, Sellers TA, Zheng W, Folsom AR. Intake of vitamins A, C, and E and postmenopausal breast cancer: The Iowa Women's Health Study. Am J Epidemiol. 1996;144(2):165–174. [PubMed]
- 145. Padayatty SJ, Riordan HD, Hewitt SM, Katz A, Hoffer LJ, Levine M. Intravenously administered vitamin C as cancer therapy: three cases. CMAJ. 2006;174:937–942. [PMC free article] [PubMed]
- 146. Kathleen A. Ascorbic acid in prevention and treatment of cancer. Altern Med Rev. 1988;3(3):174–186.
- 147. Chen Q, Espey MG, Krishna MC, Mitchell JB, Corpe CP, Buettner GR, et al. Pharmacologic ascorbic acid concentrations selectively kill cancer cells: action as a pro-drug to deliver hydrogen peroxide to tissues. Proc Natl Acad Sci USA. 2005;102:13604–13609. [PMC free article] [PubMed]
- 148. Cabanillas F. Vitamin C and cancer: what can we conclude—1, 609 patients and 33 years later? P R Health Sci J. 2010;29(3):215–217. [PubMed]
- 149. Cha J, Roomi MW, Ivanov V, Kalinovsky T, Niedzwiecki A, Rath M. Ascorbate depletion increases growth and metastasis of melanoma cells in vitamin C deficient mice. Exp Oncol. 2011;33(4):226–230. [PubMed]
- 150. Cha J, Roomi MW, Ivanov V, Kalinovsky T, Niedzwiecki A, Rath M. Ascorbate supplementation inhibits growth and metastasis of B16FO melanoma and 4T1 breast cancer cells in vitamin C-deficient mice. Int J Oncol. 2013;42(1):55–64. [PMC free article] [PubMed]

- 151. Assouline S, Miller WH. High-dose vitamin C therapy: renewed hope or false promise? CMAJ. 2006;174:956–957. [PMC free article] [PubMed]
- 152. Yeom CH, Jung GC, Song KJ. Changes of terminal cancer patients' health-related quality of life after high dose vitamin C administration. J Korean Med Sci. 2007;22:7–11. [PMC free article] [PubMed]
- 153. Wild SH, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. Diabetes Care. 2004;27:1047–1053. [PubMed] 154. American Diabetes Association Diagnosis and classification of diabetes mellitus. Diabetes Care. 2010;33:S62. [PMC free article] [PubMed]
- 155. Tesfaye S, Gill G. Chronic diabetic complications in Africa. Afr J Diabetes Med. 2011;19:4–8.
- 156. Chintan AP, Nimish LP, Nayana B, Bhavna M, Mahendra G, Hardik T. Cardiovascular complication of diabetes mellitus. J Appl Pharm Sci. 2011;4:1–6.
- 157. Vincintini Juliana MS, Valentini Juliana MS, Grotto Denise MS, Paniz C, Roehrs M, Brucker N, et al. Association among microalbuminuria and oxidative stress biomarkers in patients with type 2 diabetes. J Investig Med. 2011;59:649–654. [PubMed]
- 158. Rosen P, Nawroth PP, King G, Moller W, Tritschler HJ, Packer L. The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO MCBN, the American Diabetes Association and the German Diabetes Society. Diabetes Metab Res Rev. 2001;17:189–912. [PubMed]
- 159. Harding AH, Wareham NJ, Bingham SA, Khaw K, Luben R, Welch A, et al. Plasma vitamin C level, fruit and vegetable consumption, and the risk of new-onset type 2 diabetes mellitus: the European prospective investigation of cancer—Norfolk prospective study. Arch Intern Med. 2008;168(14):1493–1499. [PubMed]
- 160. Montenen J, Knekt P, Jarvinen R, Reunanen A. Dietary antioxidants and risk of type 2 diabetes. Diabetes Care. 2004;27:362–366. [PubMed]
- 161. Maxwell SR, Thomason H, Sandler D, Leguen C, Baxter MA, Thorpe GH, et al. Antioxidant status in patients with uncomplicated insulin-dependent and non-insulin-dependent diabetes mellitus. Eur J Clin Invest. 1997;27(6):484–490. [PubMed]
- 162. Odum EP, Ejilemele AA, Wakwe VC. Antioxidant status of type 2 diabetic patients in Port Harcourt, Nigeria. Niger J Clin Pract. 2012;15(1):55–58. [PubMed]
- 163. Mazloom Z, Hejazi N, Dabbaghmanesh MH, Tabatabaei HR, Ahmadi A, Ansar H. Effect of vitamin C supplementation on postprandial oxidative stress and lipid profile in type 2 diabetic patients. Pak J Biol Sci. 2011;14(19):900–904. [PubMed]
- 164. Sharma P, Mishra S, Ajmera P, Mathur S. Oxidative stress in metabolic syndrome. Indian J Clin Biochem. 2005;20(1):145–149. [PMC free article] [PubMed]
- 165. Özkaya D, Naziroğlu M, Armağan A, Demirel A, Köroglu BK, Çolakoğlu N, et al. Dietary vitamin C and E modulates oxidative stress induced-kidney and lens injury in diabetic aged male rats through modulating glucose homeostasis and antioxidant systems. Cell Biochem Funct. 2011;29(4):287–293. [PubMed]
- 166. Naziroğlu M, Butterworth PJ, Sonmez TT. Dietary vitamin C and E modulates antioxidant levels in blood, brain, liver, muscle, and testes in diabetic aged rats. Int J Vit Nutr Res. 2011;81(6):347–357. [PubMed]
- 167. Sridulyakul P, Wongeak-in N, Patumraj S. Correlations between endothelial functions and ROS detection in diabetic microvascular wall: early and late ascorbic acid supplementation. Int J Vasc Med. 2012;2012:709695. [PMC free article] [PubMed] 168. Hoffman RP, Dye AS, Bauer JA. Ascorbic acid blocks hyperglycaemic impairment of endothelial function in adolescents with type 1 diabetes. Pediatr Diabetes. 2012;13(8):607—
- 610. [PMC free article] [PubMed]

- 169. Prakash J, Hota J, Singh S, Sharma O. Clinical spectrum of chronic renal failure in the elderly: a hospital based study from eastern India. Int Urol Nephrol. 2006;38:821–827. [PubMed]
- 170. Qin QJ, Deng HC, Zhao TF, Cao WF, Liu DF, Lan LZ. Study on the effect and mechanism of ascorbic acid on renal podocytes in diabetes. Zhongguo Ying Yong Sheng Li Xue Za Zhi. 2008;24(1):112–115. [PubMed]
- 171. Lee EY, Lee MY, Hong SW, Chung CH, Hong SY. Blockade of oxidative stress by vitamin C ameliorates albuminuria and renal sclerosis in experimental diabetic rats. Yonsei Med J. 2007;48(5):847–855. [PMC free article] [PubMed]
- 172. Kowluru RA, Tang J, Kern TS. Abnormalities of retinal metabolism in diabetes and experimental galactosemia. VII. Effect of long-term administration of antioxidants on the development of retinopathy. Diabetes. 2001;50:1938–1942. [PubMed]
- 173. Mustata GT, Rosca M, Biemel KM, Reihl O, Smith MA, Viswanathan A. Paradoxical effects of green tea (*Camellia sinensis*) and antioxidant vitamins in diabetic rats: improved retinopathy and renal mitochondrial defects but deterioration of collagen matrix glycoxidation and cross-linking. Diabetes. 2005;4:517–526. [PubMed]
- 174. Penn JS, Madan A, Caldwell RB, Bartoli M, Caldwell RW, Hartnett ME. Vascular endothelial growth factor in eye disease. Prog Retin Eye Res. 2008;27:331–371. [PMC free article] [PubMed]
- 175. Cotter MA, Love A, Watt MJ, Cameron NE, Dines KC. Effects of natural free radical scavengers on peripheral nerve and neurovascular function in diabetic rats. Diabetologia. 1995;38(11):1285–1294. [PubMed]
- 176. Ziegler D, Sohr CG, Nourooz-Zadeh J. Oxidative stress and antioxidant defense in relation to the severity of diabetic polyneuropathy and cardiovascular autonomic neuropathy. Diabetes Care. 2004;27(9):2178–2183. [PubMed]
- 177. Singh PP, Mahadi F, Roy A, Sharma P. Reactive oxygen species, reactive nitrogen species and antioxidants in etiopathogenesis of diabetes mellitus type-2. Indian J Clin Biochem. 2009;24(4):324–342. [PMC free article] [PubMed]
- 178. Leibovitz B, Siegel BV. Ascorbic acid, neutrophil function, and the immune response. Int J Vit Nutr Res. 1978;48(2):159–164. [PubMed]
- 179. Animashaun A, Kelleher J, Heatley RV, Trejdosiewicz LK, Losowsky MS. The effect of zinc and vitamin C supplementation on the immune status of patients with Crohn's disease. Clin Nutr. 1990;9(3):137–146. [PubMed]
- 180. Maggini S, Wintergerst ES, Beveridge S, Hornig DH. Selected vitamins and trace elements support immune function by strengthening epithelial barriers and cellular and humoral immune responses. Br J Nutr. 2007;98(1):S29–S35. [PubMed]
- 181. Feigen GA, Smith BH, Dix CE, Flynn CJ, Peterson NS, Rosenberg LT, et al. Enhancement of antibody production and protection against systemic anaphylaxis by large doses of vitamin C. Res Commun Chem Pathol Pharmacol. 1982;38(2):313–333. [PubMed]
- 182. Haskell BE, Johnston CS. Complement component C1q activity and ascorbic acid nutriture in guinea pigs. Am J Clin Nutr. 1991;54(6):1228S–1230S. [PubMed]
- 183. Wintergerst ES, Maggini S, Hornig DH. Contribution of selected vitamins and trace elements to immune function. Ann Nutr Metab. 2007;51(4):301–323. [PubMed]
- 184. Jeong YJ, Hong SW, Kim JH, Jin DH, Kang JS, Lee WJ, et al. Vitamin C-treated murine bone marrow-derived dendritic cells preferentially drive naïve T cells into Th1 cells by increased IL-12 secretions. Cell Immunol. 2011;266(2):192–199. [PubMed]
- 185. Holmannová D, Koláčková M, Krejsek J. Vitamin C and its physiological role with respect to the components of the immune system. Vnitr Lek. 2012;58(10):743–749. [PubMed] 186. Mikirova N, Casciari J, Rogers A, Taylor P. Effect of high-dose intravenous vitamin C on inflammation in cancer patients. J Transl Med. 2012;10:189. [PMC free article] [PubMed]

- 187. Dwivedi S, Goel A, Mandhani A, Natu SM, Khattri S, Pant KK. Diagnostic and prognostic significance of prostate specific antigen and serum interleukin 18 and 10 in patients with locally advanced carcinoma prostate: a prospective comparative study. Asian Pacific J Cancer Prev. 2011;12:1639–1644. [PubMed]
- 188. Dwivedi S, Shukla KK, Gupta G, Sharma P. Non-invasive biomarker in prostate cancer—a novel approach. Ind J Clin Biochem. 2013;28(2):107–109. [PMC free article] [PubMed]
- 189. Khaw KT, Woodhouse P. Interrelation of vitamin C, infection, haemostatic factors and cardiovascular disease. BMJ. 1995;310:1559–1563. [PMC free article] [PubMed]
- 190. Ness AR, Powles JW, Khaw KT. Vitamin C and cardiovascular disease—a systematic review. J Cardiovasc Risk. 1997;3:513–521. [PubMed]
- 191. Woodward M, Lowe GDO, Rumley A, Tunstall-Pedoe H, Philippou H, Lane DA. Epidemiology of coagulation factors, inhibitors and activation markers: the third Glasgow MONICA study (2). Relationships to cardiovascular risk factors and prevalent cardiovascular disease. Br J Haematol. 1997;97:785–797. [PubMed]
- 192. Woodward M, Rumley A, Tunstall-Pedoe H, Lowe GDO. Association of blood rheology and interleukin-6 with cardiovascular risk factors and prevalent cardiovascular disease. Br J Haematol. 1999;104:246–257. [PubMed]
- 193. Woodward M, Rumley A, Lowe GDO, Tunstall-Pedoe H. C-reactive protein: associations with haematological variables, cardiovascular risk factors and prevalent cardiovascular disease. Br J Haematol. 2003;122:135–141. [PubMed]
- 194. Gao X, Bermudez OI, Tucker KL. Plasma C-reactive protein and homocysteine concentrations are related to frequent fruit and vegetable intake in hispanic and non-hispanic white elders. J Nutr. 2004;134:913–918. [PubMed]
- 195. Ford ES, Liu S, Mannino DM, Giles WH, Smith SJ. C-reactive protein concentration and concentrations of blood vitamins, carotenoids, and selenium among United States adults. Eur J Clin Nutr. 2003;57:1157–1163. [PubMed]
- 196. Lowe GDO. Circulating inflammatory markers and risks of cardiovascular and non-cardiovascular disease. J Thromb Haemost. 2005;3:1618–1627. [PubMed]
- 197. Wintergerst ES, Maggini S, Hornig DH. Immune-enhancing role of vitamin C and zinc and effect on clinical conditions. Ann Nutr Metab. 2006;50(2):85–94. [PubMed]
- 198. Stohs SJ, Bagchi D. Mechanisms in the toxicity of metal ions. Free Radic Biol Med. 1995;18(2):321–336. [PubMed]
- 199. Shaban El-Neweshy M, Said El-Sayed Y. Influence of vitamin C supplementation on lead-induced histopathological alterations in male rats. Exp Toxicol Pathol. 2011;63(3):221–227. [PubMed]
- 200. Kosik-Bogacka DI, Baranowska-Bosiacka I, Marchlewicz M, Kolasa A, Jakubowska K, Olszewska M, et al. Effect of l-ascorbic acid and/or tocopherol supplementation on electrophysiological parameters of the colon of rats chronically exposed to lead. Med Sci Monit. 2011;17(1):BR16–BR26. [PMC free article] [PubMed]
- 201. Houston DK, Johnson MA. Does vitamin C intake protect against lead toxicity? Nutr Rev. 2000;58(3):73–75. [PubMed]
- 202. Singh AL, Singh VK, Srivastava A. Effect of arsenic contaminated drinking water on human chromosome: a case study. 2013. doi: 10.1007/s12291-013-0330-3. [PMC free article] [PubMed]
- 203. Herrera A, Pineda J, Antonio MT. Toxic effects of perinatal arsenic exposure on the brain of developing rats and the beneficial role of natural antioxidants. Environ Toxicol Pharmacol. 2013;36(1):73–79. [PubMed]

- 204. Antonio Garcia MT, Herrera Dueñas A, Pineda Pampliega J. Hematological effects of arsenic in rats after subchronical exposure during pregnancy and lactation: the protective role of antioxidants. Exp Toxicol Pathol. 2013;65(5):609–614. [PubMed]
- 205. Singh S, Rana SV. Ascorbic acid improves mitochondrial function in liver of arsenic-treated rat. Toxicol Ind Health. 2010;26(5):265–272. [PubMed]
- 206. Singh MK, Dwivedi S, Yadav SS, Sharma P, Khattri S. Arsenic-induced hepatic toxicity and its attenuation by fruit extract of *Emblica officinalis* (amla) in mice. Ind J Clin Biochem. 2013. doi:10.1007/s12291-013-0353-9. [PMC free article] [PubMed]
- 207. El-Sokkary GH, Awadalla EA. The protective role of vitamin C against cerebral and pulmonary damage induced by cadmium chloride in male adult albino rat. Open Neuroendocrinol J. 2011;4:1–8.
- 208. Hounkpatin ASY, Johnson RC, Guédénon P, Domingo E, Alimba CG, Boko M. Protective effects of vitamin C on haematological parameters in intoxicated Wistar rats with cadmium, mercury and combined cadmium and mercury. Int Res J Biol Sci. 2012;1(8):76–81. 209. Gajawat S, Sancheti G, Goyal PK. Vitamin C against concomitant exposure to heavy
- metal and radiation: a study on variations in hepatic cellular counts. Asian J Exp Sci. 2005;19(2):53–58.
- 210. Lucksch F. C-vitamin und schizophrenic. Wien Klin Wochenschr. 1940;53:1009–1011.
- 211. Subotičanec K, Folnegović-Šmalc V, Korbar M, Meštrović B, Buzina R. Vitamin C status in chronic schizophrenia. Biol Psychiatry. 1990;28(11):959–966. [PubMed]
- 212. Altuntas I, Aksoy H, Coskun I, Caykoylu A, Akcay F. Erythrocyte superoxide dismutase and glutathion peroxidase activities and malondialdehyde and reduced glutathione levels in schizophrenic patients. Clin Chem Lab Med. 2000;38:1277–1281. [PubMed]
- 213. Vaiva G, Thomas P, Leroux JM. Erythrocyte superoxide dismutase determination in positive moments of psychosis. Therapie. 1994;49:343–348. [PubMed]
- 214. Reddy R, Mahadik SP, Mukherjee M, Murty JN. Enzymes of the antioxidant system in chronic schizophrenic patients. Biol Psychiatry. 1991;30:409–412. [PubMed]
- 215. Kuloghi M, Ustundag B, Atmaca M, Canatan H, Tezean AE, Cinkiline N. Lipid peroxidation and antioxidant enzyme levels in patients with schizophrenia and bipolar disorder. Cell Biochem Funct. 2002;20:171–175. [PubMed]
- 216. Mahadik SP, Mukherjee S, Correnti I, Sheffer R. Elevated levels of lipid peroxidation products in plasma from drug-naive patients at onset of psychosis. Schizophr Res. 1995;15:66–70.
- 217. Arvindakshan M, Sitasawad S, Debsikdar V, Ghate M, Evans D, Horrobin DF, et al. Essential polyunsaturated fatty acid and lipid peroxide levels in never-medicated and medicated schizophrenia patients. Biol Psychiatry. 2003;53:56–64. [PubMed]
- 218. Gaur N, Gautam S, Gaur M, Sharma P, Dadheech G, Mishra S. The biochemical womb of schizophrenia: a review. Indian J Clin Biochem. 2008;23(4):307–327. [PMC free article] [PubMed]
- 219. Fridovich I. Superoxide radical: an endogenous toxicant. Annu Rev Pharmacol Toxicol. 1983;23:239–257. [PubMed]
- 220. Dadheech G, Mishra S, Gautam S, Sharma P. Evaluation of antioxidant deficit in schizophrenia. Indian J Psychiatry. 2008;50(1):16–20. [PMC free article] [PubMed]
- 221. Seregi A, Schaefer A, Komlos M. Protective role of brain ascorbic acid content against lipid peroxidation. Experientia. 1978;34:1056–1057. [PubMed]
- 222. Oke AI, May L, Adams RN (1987). Ascorbic acid distribution pattern in human brain. Ann NY Acad Sci. 1987;498:1–12. [PubMed]
- 223. Arvindakshan M, Ghate M, Ranjekar PK, Evans DR, Mahadik SP. Supplementation with a combination of omega 3 fatty acids and antioxidants (vitamin E and C) improves the outcome of schizophrenia. Schizophr Res. 2003;62:195–204. [PubMed]

- 224. Dadheech G, Mishra S, Gautam S, Sharma P. Oxidative stress—tocopherol, ascorbic acid and reduced glutathione status in schizophrenics. Indian J Clin Biochem. 2006;21(2):34–38. [PMC free article] [PubMed]
- 225. Dakhale GN, Khanzode SD, Khanzode SS, Saoji A. Supplementation of vitamin C with atypical antipsychotics reduces oxidative stress and improves the outcome of schizophrenia. Psychopharmacology. 2005;182:494–498. [PubMed]
- 226. Gautam M, Agrawal M, Gautam M, Sharma P, Gautam AS, Gautam S. Role of antioxidants in generalised anxiety disorder and depression. Indian J Psychiatry. 2012;54(3):244–247. [PMC free article] [PubMed]
- 227. Harrison FE. A critical review of vitamin C for the prevention of age-related cognitive decline and Alzheimer's disease. J Alzheimers Dis. 2012;29(4):711–726. [PMC free article] [PubMed]

Articles from Indian Journal of Clinical Biochemistry are provided here courtesy of **Springer**